Phenomenological Incorporation of Nonlinear Dendritic Integration Using Integrate-and-Fire Neuronal Frameworks

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Abstract

It has been discovered recently in experiments that the dendritic integration of excitatory glutamatergic inputs and inhibitory GABAergic inputs in hippocampus CA1 pyramidal neurons obeys a simple arithmetic rule as $V_S^{\text{Exp}} \approx V_E^{\text{Exp}} + V_I^{\text{Exp}} + k V_E^{\text{Exp}} V_I^{\text{Exp}}$, where V_S^{Exp} , V_E^{Exp} and V_I^{Exp} are the respective voltage values of the summed somatic potential, the excitatory postsynaptic potential (EPSP) and the inhibitory postsynaptic potential measured at the time when the EPSP reaches its peak value. Moreover, the shunting coefficient k in this rule only depends on the spatial location but not the amplitude of the excitatory or inhibitory input on the dendrite. In this work, we address the theoretical issue of how much the above dendritic integration rule can be accounted for using subthreshold membrane potential dynamics in the soma as characterized by the conductance-based integrate-and-fire (I&F) model. Then, we propose a simple I&F neuron model that incorporates the spatial dependence of the shunting coefficient k by a phenomenological parametrization. Our analytical and numerical results show that this dendritic-integration-rule-based I&F (DIF) model is able to capture many experimental observations and it also yields predictions that can be used to verify the validity of the DIF model experimentally. In addition, the DIF model incorporates the dendritic integration effects dynamically and is applicable to more general situations than those in experiments in which excitatory and inhibitory inputs occur simultaneously in time. Finally, we generalize the DIF neuronal model to incorporate multiple inputs and obtain a similar dendritic integration rule that is consistent with the results obtained by using a realistic neuronal model with multiple compartments. This generalized DIF model can potentially be used to study network dynamics that may involve effects arising from dendritic integrations.

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Introduction

A single neuron may receive and integrate thousands of excitatory and inhibitory synaptic inputs from its dendritic tree. Through spatial and temporal integration of these synaptic inputs, neurons in the cortex process information efficiently and produce output signals known as spike trains. In order to understand how the brain works, it is important to understand the rules that govern dendritic integration. However these rules remain to be fully elucidated. For the integration of multiple excitatory inputs, cable theory [1,2] was developed and was successfully applied to describe passive properties of dendrites as observed in experiments [3,4]. In order to consider active properties of dendrites such as the activity of some voltage-gated ion channels and the occurrence of dendritic spikes [5-7], a two-layer network model was proposed [8,9]. This model has been supported by some experiments using focal synaptic stimulation and glutamate uncaging [10,11]. However, as reviewed in Refs. [12,13], there are still many other

important properties of dendritic integration, such as spike timing, that cannot be captured by this model.

In contrast to theoretical and experimental results for the integration rule of multiple excitatory inputs as mentioned above, we know even less about the integration rule when both excitatory and inhibitory inputs are presented together. In order to describe dynamics of neuronal circuitry within the brain, it is important to understand how excitatory and inhibitory inputs are integrated. For example, the interactions of excitatory and inhibitory synaptic currents have been found to play an important role in many sensory systems [14–16]. Shunting inhibition is found to be able to control the gain of a neuron in the presence of excitatory synaptic inputs [17]. Inhibition can also modulate the frequency [18] and improve the robustness [19] of gamma oscillations through nonlinear interactions with synaptic excitation. Therefore, it is important to determine precise rules that govern synaptic excitation-inhibition integration in order to achieve a good

understanding of underlying computational mechanisms for these neurophysiological phenomena.

Recently, a quantitative description of a dendritic integration rule has been uncovered in experimental results from CA1 pyramidal neurons in one of our authors' lab [20]. In the experiment, when the excitatory glutamatergic input and the inhibitory GABAergic input were elicited simultaneously with two iontophoretic pipettes at adjacent locations on a dendritic trunk, the response measured in the soma was found to be always smaller than the linear sum of the individual excitatory postsynaptic potential (EPSP) and inhibitory postsynaptic potential (IPSP) measured in the soma separately as shown in Fig. 1. In Fig. 1, t^* denotes the time when the EPSP reaches its peak value, denoted as V_F^{Exp} , which is referred to as the amplitude of EPSP. The values of IPSP and the summed somatic potential (SSP) at time t^* were denoted by V_I^{Exp} and V_S^{Exp} , referred to as the amplitudes of IPSP and SSP, respectively. The arithmetic summation rule for the dendritic integration in Ref. [20] can now be expressed as

$$V_{\rm S}^{\rm Exp} \approx V_E^{\rm Exp} + V_{\rm I}^{\rm Exp} + k V_E^{\rm Exp} V_I^{\rm Exp},\tag{1}$$

where the third term on the right-hand side of Eq. (1) is the socalled shunting component (SC) with k as the shunting coefficient [20]. Such a relationship was also found for the mean values of EPSP, IPSP and SSP, instead of the voltage values at time t^* [20]. In addition, the shunting coefficient k depends on locations of both the excitatory and the inhibitory stimulus but not the amplitudes of EPSP and IPSP. As shown in the inset of Fig. 2, when the location of the inhibitory input on the dendritic trunk is fixed and the excitatory input is located in between the soma and the inhibitory input site, k increases as the distance between the excitatory input and the soma increases. On the other hand, when the excitatory input is located further away from the soma than the inhibitory input site, k remains almost constant with further increases in the distance between the excitatory input site and the soma.

In Ref. [20], numerical simulation based on the software NEURON [21] was also performed to examine this integration rule. The simulation used a reconstructed spatial structure of the CA1 neuron, which included 200 compartments, four different ion channels, and four different neurotransmitter receptors. These components are used to mimic both the active channel properties and passive cable properties of the dendrite of the real neuron. The simulation was able to account for many aspects of the experimental results. However, the value of k produced by the simulation was approximately only one half of the experimentally measured value, indicating that the constructed multi-compartmental neuron is still not able to capture quantitatively the effects of dendritic integration of a real neuron. Moreover, there are some other experimental results as reported in the supplementary text of Ref. [20] that have not been addressed by this multi-compartmental model, such as the case when excitatory inputs and inhibitory inputs are no longer synchronous--clearly, this situation often occurs and excitatory and inhibitory stimulations may not always occur at precisely exactly the same time as in Ref. [20]. (Of course, excitatory and inhibitory events can be often highly correlated in time [22].) We further note that the model used in simulations contains many compartments and parameters, rendering it difficult to study analytically. In addition, there are other theoretical issues that need to be clarified. For example, it is not clear how the nonlinear SC term arises mechanistically by using this simulation approach. Furthermore, neuronal coding often involves dynamics of networks. However, it would be difficult to implement this multi-compartmental model in network simulation



Figure 1. Experimental measurement of EPSP, IPSP and SSP. The time when EPSP reaches its peak value is denoted by t^* . V_E^{Exp} and V_I^{Exp} represent the amplitude of EPSP and IPSP at time t^* , respectively. V_S^{Exp} represents the amplitude of SSP at time t^* . SC is the difference between the SSP and the linear sum of the individual EPSP and IPSP measured under separate excitatory and inhibitory inputs. When the excitatory input and the inhibitory input are elicited simultaneously, the response amplitude measured at the soma is found to be smaller than that of the linear sum. (modified from Ref. [20]). doi:10.1371/journal.pone.0053508.g001

in that the complexity of such a neuron model would make the computational cost impractical. Therefore, it is desirable to incorporate the dendritic integration features into a simpler neuronal model that has the potential to address these theoretical issues.

Many spiking neuronal models have been developed to capture spike dynamics of real neurons [23-30]. Each model has its own advantages and disadvantages with respect to the understanding of neuronal spiking dynamics [31]. Since the data measured in the experiment [20] were all collected at the soma, we first address the issue of whether we can understand this dendritic integration rule through somatic properties of a single-compartment point-neuron model. Considering the trade-off between biological plausibility and theoretical complexity among those existing neuronal models [31-34], we choose the conductance-based integrate-and-fire (I&F) model as the basic model for the investigation of the dendritic integration rule. Note that the voltage traces measured in the experiment [20] involved only subthreshold dynamics. The I&F model is well-suited to investigate the dendritic integration rule as models of the I&F type have been experimentally shown to quantitatively capture the subthreshold dynamics of neurons [34,35]. Surprisingly, this simple model can produce the arithmetic rule [Eq. (1)] for the case in which the stimulus location is *fixed*. Therefore, it suggests that somatic membrane potential dynamics may play a role in the so-called dendritic integration rule. Through our theoretical analysis, we demonstrate that the nonlinear SC term arises from the multiplication of conductance and voltage in the synaptic input of the neuron. We further point out that, in a static two-port analysis, the product between excitatory and inhibitory conductances gives rise to the product between EPSP and IPSP. Using the insight derived from these analyses, finally, we develop a dendritic-integration-rule-based I&F (DIF) model to phenomenologically incorporate the spatial





Figure 2. The spatial dependence of $-\alpha$ **obtained from Eq. (9).** Given three fixed inhibitory input locations, we can parameterize α in Eq. (9) by identifying k^M with the shunting coefficient measured in the experiment [20] to obtain the spatial dependence of α . Inset: the experimental measurements of the shunting coefficient [20] as a function of the distance between the excitatory input site and the soma for three fixed inhibitory input sites on the dendrite (marked by dashed lines).

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effect of the integration. This phenomenological model can not only incorporate the dendritic integration rule, but also capture experimental observations that have not been addressed by the multi-compartmental model [20]. In addition, it also yields predictions that can be further used to validate this model experimentally. We note that the original dendritic integration rule [Eq. (1)] is limited to the situation in which the excitatory and inhibitory inputs are concurrent. When excitatory and inhibitory inputs are not concurrent, the shunting coefficient k has to be measured again in experiments to obtain the corresponding dendritic integration formula. However, we show that the DIF model has extended the dendritic integration rule to dynamical situations and it can be used to study neuronal responses to any stimulus pattern with both excitatory and inhibitory inputs. Finally, we generalize the DIF model for studying the effect of multiple inputs and obtain related results that are consistent with those obtained using the NEURON software as reported in Ref. [20]. Because our model is an I&F type, clearly, it can easily be implemented in network dynamics to study certain effects arising from dendritic integrations.

The paper is organized as follows, we first show both theoretically and numerically that the arithmetic dendritic integration rule can partially be explained by somatic subthreshold membrane potential dynamics of the conductance-based I&F model. Next, using the insight derived from a static two-port analysis, we develop a simple DIF model to phenomenologically parameterize the spatial dependence of the shunting coefficient k. Our numerical results show that this model captures many neurophysiological phenomena as observed in experiments. Here, we further extend the DIF model to account for multiple inputs. In *Discussion*, we determine the parameter range and describe some predictions of the DIF model. In *Methods*, we introduce the conductance-based I&F model and present both analytical and



Figure 3. Reproduced profiles of EPSP and IPSP by the l&F model. The thick dark (blue online) lines are produced from the l&F model, the light gray lines represent EPSP and IPSP measured in the experiment for different trials, and the thin dark (red online) lines represent the trial-averaged responses in the experiment. Parameters in the I&F model are chosen as follows, $f_E = 1.16 \times 10^{-5} S \cdot cm^{-2}$, $\sigma_{Er} = 5 ms$, $\sigma_{Ed} = 7.8 ms$, $f_I = 3.71 \times 10^{-5} S \cdot cm^{-2}$, $\sigma_{Ir} = 6 ms$, and $\sigma_{Id} = 18 ms$. (See *Methods* for details). doi:10.1371/journal.pone.0053508.0003

approximate solutions to the I&F-type models. Here, we also recapitulate the static two-port analysis in detail.

Results

Subthreshold Membrane Potential in the Soma

It has been demonstrated that the I&F model can capture very well the subthreshold membrane potential dynamics in the soma of a real neuron when its membrane potential is below ~ -55 mV [34,35]. The membrane potential used in the experiment [20] for determining the dendritic integration rule is precisely in this subthreshold regime. A natural question arises, that is, how much the dendritic integration rule can be accounted for by the somatic membrane dynamics. Here, we employ the conductance-based I&F model (see Methods) to address this question. First, we use the experimental data to determine appropriate parameters to reproduce the same profiles of EPSP and IPSP as measured in the experiment [20] (Fig. 3, see Methods for details). Then, we use a fourth-order Runge-Kutta method to solve the I&F model numerically. We denote the EPSP by $V_E(t)$, IPSP by $V_I(t)$, and SSP by $V_S(t)$. The time t^* is the time when $V_E(t)$ reaches its peak value. The values of $V_E(t^*)$, $V_I(t^*)$ and $V_S(t^*)$ are referred to as the amplitudes of EPSP, IPSP and SSP, respectively. In order to verify the product form of the SC term, we follow the same data processing procedure as in Ref. [20]: by setting the EPSP amplitude at a fixed value while varying the IPSP amplitude, we examine whether the SC amplitude linearly depends on the IPSP amplitude. Conversely, for a fixed amplitude of IPSP, we examine whether the SC amplitude linearly depends on the EPSP amplitude. As shown in Fig. 4A, the SC increases linearly with respect to the IPSP amplitude when the EPSP amplitude is fixed, whereas the SC increases linearly with respect to the EPSP amplitude when the IPSP amplitude is fixed. In addition, we observe that both straight lines in Fig. 4A have almost identical slopes and this relationship is exactly the same as the one observed in the experiment [20]. If we use the mean values \bar{V}_E , \bar{V}_I and \bar{V}_S of EPSP, IPSP and SSP respectively, averaged over a time interval



Figure 4. Dendritic integration rule obtained from the l&F model (A–C) and the modified l&F model (D–F). We choose the values of $\alpha = -8k\Omega \cdot \text{cm}^2$ and $\beta = 7k\Omega \cdot \text{cm}^2$ so that the value of k computed from Eq. (8) matches the value measured in the experiment [20]. Lines in figures indicate linear fits with slope k. (A) Ratio between SC and EPSP (SC/EPSP) plotted against IPSP (red square online) and SC/IPSP plotted against EPSP (blue circle online) at time t^* for the l&F model (Red online: k = 0.070; blue online: k = 0.065). Inset: experimental measurement (Red online: k = 0.142; blue online: k = 0.145). (B) Ratio between the mean SC and the mean EPSP (SC/EPSP) plotted against the mean IPSP (red square online), the mean SC and the mean IPSP (SC/IPSP) plotted against the mean IPSP (red square online), the mean SC and the mean IPSP (SC/IPSP) plotted against the mean IPSP (sc/IPSP) plotted against the mean SC and the mean EPSP (blue circle online) for the l&F model. Inset: experimental measurement. (C) the same as (A) but at time (1) t = 20 ms, (2) t = 40 ms, (3) t = 60 ms and (4) t = 80 ms, respectively, as marked on the figures. (D) the same as (A) but for the modified l&F model. (Red online: k = 0.143). Inset: the same as the inset of (A). (E) the same as (C) but for the modified l&F model. Inset: the same as the inset of (B). doi:10.1371/journal.pone.0053508.q004

of 100 ms, the summation rule $\bar{V}_S = \bar{V}_E + \bar{V}_I + \bar{k}\bar{V}_E\bar{V}_I$ also holds as shown in Fig. 4B. This is also consistent with the experimental observations [20]. Our numerical results further show that the rule holds for $V_E(t)$, $V_I(t)$ and $V_S(t)$ at any moment of time *t* (i.e., not restricted to t^*) as demonstrated in Fig. 4C (1–4) for a selected set of times.

As shown above, the I&F dynamics of somatic membrane potential can exhibit the dendritic integration rule [Eq. (1)]. However, one may ask why this linear dynamics of somatic subthreshold membrane potential as described by the I&F model can produce such a nonlinear integration rule. Below, we answer this question analytically (see *Methods* for details). From Eq. (1), we have

$$k = \frac{V_S(t) - V_E(t) - V_I(t)}{V_E(t) \cdot V_I(t)}.$$
 (2)

If we can demonstrate that k is independent of the amplitude of inputs, then the dendritic integration rule holds. Performing a Taylor expansion of the analytical solution to the conductancebased I&F model, we can obtain approximations of EPSP, IPSP and SSP (See *Methods*). Substituting these voltage values [Eqs. (22b), (23b) and (24b)] into Eq. (2), we can obtain an approximate k, denoted by k^* as

$$k^{*} = -\frac{\int_{0}^{t} \{\varepsilon_{E} h_{I}(u) Q(h_{E}, u) + \varepsilon_{I} h_{E}(u) Q(h_{I}, u)\} du}{\varepsilon_{E} \varepsilon_{I} Q(h_{E}, t) Q(h_{I}, t)}, \qquad (3)$$

where Q(f,x) is a functional defined as $Q(f,x) = \int_0^x e^{G_L(y-t)/C} f(y) dy$. ε_E and ε_I are the excitatory and inhibitory reversal potentials, respectively. $h_E(\cdot)$ and $h_I(\cdot)$ as given by Eqs. (17) and (18) only determine the profiles (rise and decay time scale) of EPSP and IPSP, whereas the excitatory and inhibitory inputs strength f_E and f_I corresponding to the amplitudes of EPSP and IPSP as in Eqs. (15) and (16), do not appear in the expression of k^* [Eq. (3)]. Therefore, k^* is independent of the amplitudes of EPSP and IPSP and IPSP.

For the mean EPSP and the mean IPSP, we can simply take integrals of V_E , V_I , V_S over the time interval [0,T] to obtain the mean values (T = 100ms in the experiment [20]), where the mean is defined as $\frac{1}{T} \int_0^T V(t) dt$. By definition, the shunting coefficient for the mean case can be evaluated as

$$k_{\text{mean}}^* = -\frac{T\int_0^T\int_0^t \{\varepsilon_E h_I(u)Q(h_E,u) + \varepsilon_I h_E(u)Q(h_I,u)\} dudt}{\varepsilon_E \varepsilon_I\int_0^T Q(h_I,t)dt\int_0^T Q(h_E,t)dt}, \quad (4)$$

which shows that k_{mean}^* is independent of the EPSP and IPSP amplitudes for the same reason as mentioned in the analysis of Eq. (3). Therefore, the dendritic integration rule holds for the somatic membrane potential as modeled by the I&F dynamics. Through

the above analysis, it can be seen that the nonlinearity in the dentritic integration rule ultimately arises from the product term of the conductance G and the voltage V in the input current of the I&F neuron (see *Methods*).

The DIF Neuronal Model

As is well known, the conductance-based I&F model is used to describe the membrane potential dynamics in the soma without taking into account dendritic structures. From the above analysis, it seems that the subthreshold membrane potential dynamics in the soma is able to explain the dendritic integration rule [Eq. (1)]. However, as can be seen in Fig. 4A, there is a difference of a factor of two between the value of the shunting coefficient k measured in the experiment and that of k computed by the I&F model. Incidentally, the experimentally measured k and the value obtained by the NEURON software in Ref. [20] also differ almost by a factor of two. Importantly, as observed in the experiment [20], the value of k depends on the distance between the input sites and the soma. However, from Eq. (3), it can be seen that k does not contain any spatial information explicitly. In the following, we will construct a simple phenomenological neuron model that incorporates the spatial dependence in the dendritic integration rule. As can be seen below, our new neuron model can account for additional observed experimental phenomena and will also be useful for network simulations that take into account effects arising from dendritic integrations.



Figure 5. The shunting coefficient k^M **as a function of time.** With three different values of β : $\beta = 10k\Omega \cdot cm^2$ (thick dark line: red online), $\beta = 0k\Omega \cdot cm^2$ (thick gray line: blue online), $\beta = -10k\Omega \cdot cm^2$ (light gray line: green online). Here α is fixed as $\alpha = -8k\Omega \cdot cm^2$. It can be seen that k^M is nearly independent of the parameter β . doi:10.1371/journal.pone.0053508.g005

To incorporate the spatial dependence of the dendritic integration rule, we first motivate the construction of our neuron model by a static two-port analysis [33] (see Methods for details). For the time-independent case, the conductance inputs of both excitation G_E and inhibition G_I are constant, the transfer resistance between any two sites on the dendritic tree is also constant. Therefore, we can express the excitatory and inhibitory currents as $I_{ex} = G_E(\varepsilon_E - V_E^E)$ and $I_{in} = G_I(\varepsilon_I - V_I^I)$, respectively. Here V_E^E represents the membrane potential at the site of excitation and V_I^I the membrane potential at the site of inhibition. We can obtain the membrane potentials V_E^E and V_I^I as $V_E^E = K_{EE}I_{ex} + K_{IE}I_{in}$ and $V_I^I = K_{II}I_{in} + K_{EI}I_{ex}$, respectively [33]. Here K_{EE} and K_{II} are local transfer resistances at the sites of excitation and inhibition, respectively. K_{IE} corresponds to the transfer resistance from the inhibitory site to the excitatory site and vice versa for K_{EI} . The membrane potential at the soma can be obtained by adding the excitatory and inhibitory contributions, i.e., $V_S = K_{ES}I_{ex} + K_{IS}I_{in}$. If the conductance inputs are sufficiently small, we can obtain the following relationship under the simultaneous drive

$$V_S \approx V_E + V_I - (K_{ES} K_{IE} \varepsilon_I + K_{IS} K_{EI} \varepsilon_E) G_E G_I, \qquad (5)$$

where $V_E(V_I)$ is the EPSP (IPSP) obtained with only excitatory (inhibitory) input, and V_S is the SSP in this static case. From *Methods*, we have that the product of V_E and V_I can be approximated by

$$V_E V_I \approx K_{ES} K_{IS} \varepsilon_E \varepsilon_I G_E G_I. \tag{6}$$

Therefore, we can obtain Eq. (39), which has exactly the same form as the dendritic integration rule with a shunting coefficient $k = -\frac{K_{IE}}{K_{IS}\varepsilon_E} - \frac{K_{EI}}{K_{ES}\varepsilon_I}$. We emphasize that this dendritic integration rule obtained through the two-port analysis is only valid for the static case. Clearly, we need to address the time-dependent case as in the experimental setup [20]. However, this static analysis provides us with an observation about possible mechanisms underlying the dendritic integration rule. Note that, in Eqs. (5) and (6), the shunting component encompasses the product term between the excitatory conductance G_E and the inhibitory conductance G_I , in turn, yielding the product, $V_E V_I$, in Eq. (39). Therefore, the product, $G_E G_I$, is the intrinsic origin of the shunting component in the static case [Eq. (38)]. Using this observation, we propose to generalize the I&F dynamics by introducing terms of $G_E G_I$ and obtain the following governing equation for a neuron:

$$C\frac{dV^{M}}{dt} = -G_{L}(V^{M} - \varepsilon_{L}) - G_{E}(1 + \alpha G_{I})(V^{M} - \varepsilon_{E}) - G_{I}(1 + \beta G_{E})(V^{M} - \varepsilon_{I}),$$
(7)

where *C* is the membrane capacitance, G_L is the leaky conductance, G_E and G_I are the excitatory and inhibitory conductances, respectively. ε_L is the resting potential, ε_E and ε_I are the excitatory and inhibitory reversal potentials, respectively. In Eq. (7), for clarity of later discussions, we denote the membrane potential by V^M . The parameters α and β are used to parameterize the spatial effects of dendritic integration. In the following, we will refer to Eq. (7) as the modified I&F model. First, we discuss how to determine the parametrization of α and β . Considering the time-independent case, we denote individual EPSP and IPSP at the soma by V_E^M and V_I^M , the SSP by V_S^M . The condition $dV^M/dt = 0$ leads to $V_E^M = \frac{G_E \varepsilon_E}{G_E + G_L}$, $V_I^M = \frac{G_I \varepsilon_I}{G_I + G_L}$ and $V_S^M = \frac{G_E \varepsilon_E + G_I \varepsilon_I + (\alpha \varepsilon_E + \beta \varepsilon_I) G_E G_I}{G_E + G_I + (\alpha + \beta) G_E G_I + G_L}$. By imposing the condition that the dendritic integration rule holds, i.e., $V_S^M = V_E^M + V_I^M + kV_E^M V_I^M$, we can conclude that the parameters α and β must satisfy $\alpha + \beta = \frac{1}{G_L}$ and $\alpha \varepsilon_E + \beta \varepsilon_I = \frac{\varepsilon_E + \varepsilon_I + k\varepsilon_E \varepsilon_I}{G_L}$, from which, we obtain $\alpha = \frac{\varepsilon_E + k\varepsilon_E \varepsilon_I}{G_L(\varepsilon_E - \varepsilon_I)}$ and $\beta = \frac{1}{G_L} - \frac{\varepsilon_E + k\varepsilon_E \varepsilon_I}{G_L(\varepsilon_E - \varepsilon_I)}$. Note that, in the limit of $G_L \to \infty$, we have $\alpha = \beta = 0$, and the modified I&F model reduces to the standard I&F model. That is, the modified I&F model is naturally a generalized I&F model. We will show below that α , β can indeed

dynamical situation. We now turn to the discussion that in the modified I&F model the dendritic integration rule holds for any moment of time, including the time when the EPSP reaches its peak. Solving the modified I&F model numerically using the fourth-order Runge-Kutta method, we have the relationship among V_E^M , V_I^M , and V_S^M as Eq. (1) at the time when $V_E^M(t)$ reaches its peak (Fig. 4D) as well as at any other times (Fig. 4E(1–4)). As shown in Fig. 4F, this integration rule is also valid if the mean values of EPSP, IPSP and SSP are used.

be used to parameterize the dendritic integration rule in a

We can further obtain a theoretical expression of the shunting coefficient. Performing Taylor expansion to the solution of integral form of the modified I&F model to obtain the approximations of EPSP, IPSP, and SSP (see *Methods* for details), then using Eq. (2), we arrive at the shunting coefficient k^M for the modified I&F model

$$k^{M} = k^{*} + \frac{(\alpha \varepsilon_{E} + \beta \varepsilon_{I})C \int_{0}^{t} h_{E}(u)h_{I}(u)e^{G_{L}(u-t)/C}du}{\varepsilon_{E}\varepsilon_{I}Q(h_{I},t)Q(h_{E},t)}, \quad (8)$$

where k^* is the shunting coefficient computed in the original I&F model [Eq. (3)]. $Q(\cdot, t)$ is the same functional as defined in Eq. (3). In Eq. (8), the first term k^* on the right-hand side is independent of the amplitude of EPSP or IPSP, as was shown previously. For the second term, similarly, because $h_E(\cdot)$ and $h_I(\cdot)$ only control the profiles of EPSP and IPSP and are independent of the amplitudes of EPSP and IPSP. Therefore, k^M is independent of the amplitudes of EPSP and IPSP and the dendritic integration rule holds for the modified I&F model. Following the same procedure, we can also show that this integration rule holds for the mean potentials. We can further approximate the shunting coefficient k^M as

$$k^{M} \approx k^{*} + \frac{\alpha C \int_{0}^{t} h_{E}(u) h_{I}(u) e^{G_{L}(u-t)/C} du}{\varepsilon_{I} Q(h_{E}, t) Q(h_{I}, t)},$$
(9)

in which k^M is independent of the parameter β . This independence is a consequence of the fact that the magnitude of the inhibitory reversal potential ($\varepsilon_I = -10 \text{ mV}$) can be viewed as much smaller than that of the excitatory one ($\varepsilon_E = 70 \text{ mV}$) in absolute value. Eq. (9) as an approximation for k^M has been verified numerically: Fig. 5 shows that k^M is indeed nearly





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Figure 6. Comparison of DIF model with experiments (A–C), multiple inputs (D), Predictions (E), spatial dependence of k in the l&F model (F). (A) SSP (measured sum) vs. the linear sum between EPSP and direct somatic hyperpolarization. Here, SC is not observed. The inhibition is caused by direct injection of an inhibitory current with amplitude of -2.6 mV (red circle online) and of -1.3 mV (red square online). Inset: experimental measurement (modified from Ref. [20]). (B) SC vs. the relative time delay between IPSP and EPSP. For fixed inhibitory input site, we choose two different input sites for excitation: one corresponds to the distal dendrite (red circle online) and the other the proximal dendrite (blue square online). Inset: experimental measurement (modified from Ref. [20]). (C) SC is not affected by changing the driving force (DF) of IPSP from 0 to -10 mV. Inset: experimental measurement (modified from Ref. [20]). (D) Dendritic integration rule for two excitatory and two inhibitory inputs. The simulated E–I sum represents the SSP obtained from Eq. (12) with a coincident activation of all excitatory and inhibitory inputs, whereas the predicted E–I sum represents the somatic membrane potential obtained from Eq. (11). (E) Comparison between k^M and k^* as a function of time. The remainder term is defined as $k^M - k^*$ (thick gray, blue online). Inset: The asymptotic behavior $\sim t^{-1}$ (denoted by "asymptotic", thick dark dash line, red online) of the remainder term (denoted by "exact", thick gray, blue online) obtained from Eq. (9). (F) spatial dependence of k^* in the l&F model when the distance increases between the excitatory input site and the soma. This is not consistent with the experimental observation (Inset: the same as the inset of Fig. 2).

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independent of β . Therefore, we can set β to be zero to obtain a further simplified form of the modified I&F model,

$$C\frac{dV^{M}}{dt} = -G_{L}(V^{M} - \varepsilon_{L}) - G_{E}(1 + \alpha G_{I})(V^{M} - \varepsilon_{E}) - G_{I}(V^{M} - \varepsilon_{I}).$$
(10)

Eq. (10) is our central result and we will refer to this model as a dendritic-integration-rule-based I&F (DIF) model. First, we comment that the above conclusions about the dendritic integration rule remain unchanged for the DIF model. Second, we show that we can incorporate the spatial dependence of the dendritic integration rule using the following parametrization. For a given pair of excitatory and inhibitory inputs, the shunting coefficient k can be measured in the experiment. We can use this measured k as the value of k^M to determine the value of α from Eq. (9), namely, we can phenomenologically fit the value of α as a function of stimulus locations. Our fitting yields the results which are shown in Fig. 2. The parameter α captures the integration effects to the soma arising from both passive cable properties and active conductance properties (ion channels and receptors) of the dendrite [20]. We note that α is determined by the shunting coefficient k measured with a pair of *concurrent* excitatory and inhibitory inputs. However, even when excitatory and inhibitory inputs are not concurrent, the shunting coefficient k obtained using Eq. (9) is still independent of the amplitudes of EPSP and IPSP because the amplitudes of EPSP and IPSP do not appear in the numerator and denominator. Therefore, the DIF model can be used to study neuronal responses to general stimulus patterns of excitatory and inhibitory inputs.

Finally, we show that the DIF model is consistent with many other experimental observations, some of which have not been obtained by the multi-compartmental model using NEURON software [20].

(1) It has been found in the experiment that SC vanishes when hyperpolarization is induced by somatic current injection $I_{inj}(t)$ instead of conductance input $G_I(t)$ on the dendrite [20]. For this case, the drive can be modeled by $I_{inj} = f \frac{\sigma_r \sigma_d}{\sigma_d - \sigma_r} \left(e^{-\frac{t}{\sigma_d}} - e^{-\frac{t}{\sigma_r}} \right)$, where f is the magnitude, σ_r and σ_d are the rise and decay time constants, respectively [33]. The dynamics of the DIF model becomes

$$C\frac{dV^{M}}{dt} = -G_{L}(V^{M} - \varepsilon_{L}) - G_{E}(V^{M} - \varepsilon_{E}) + I_{inj}.$$

As verified numerically in *Methods*, the soma response V_E^M under only excitatory drive can be well approximated by the first-order expansion as $V_E^M \approx \int_0^t \varepsilon_E \frac{G_E(u)}{C} e^{G_L(u-t)/C} du$. Therefore, for multiple excitatory inputs, we have the linear summation of EPSPs. This linear summation rule for EPSP has also been found in experiments [36]. We can further obtain $V_S^M \approx V_E^M + V_I^M$: Fig. 6A reproduces the experimental observation (the inset of Fig. 6A) that there is no longer a nonlinear SC term.

(2) It has also been examined how the amplitude of SC is affected by a relative temporal delay between excitatory and inhibitory inputs. In the experiment [20], it was found that (i) the SC amplitude decreases with the length of delay interval between excitation and inhibition and (ii) A larger SC is induced when the excitatory input is located on the distal dendrite than that on the proximal dendrite. The experimental observation (i) can be explained as follows. The shorter the temporal delay between excitation and inhibition, the larger the SC because the amplitude of SC relies on the product between $V_E(t)$ and $V_I(t)$. Due to this product form, there is a kink structure for the SC, which can be seen both in the experiment (the inset of Fig. 6B) and in our DIF model (Fig. 6B). The observation (ii) can be understood as follows: A distal excitatory input indicates a larger shunting coefficient k, which leads to a larger absolute value of α in Eq. (9), therefore, SC is larger for the distal excitatory input. Indeed, the DIF model can capture this time-delay shunting effect successfully as shown in Fig. 6B. The above experimental phenomena have not been addressed by the multi-compartmental model in Ref. [20].

(3) By changing the driving force for IPSP from -10 to 0 mV, it has been found in the experiment [20] that the nonlinear SC term is not affected (the inset of Fig. 6C). In the DIF model, SC can be obtained as $SC = -\int_0^t \{\varepsilon_E \frac{G_I(u)}{C^2} Q(G_E, u) + \varepsilon_I \frac{G_E(u)}{C^2} Q(G_I, u) - (\frac{\alpha}{C} \varepsilon_E + \frac{\beta}{C} \varepsilon_I) G_E(u) G_I(u) e^{G_L(u-t)/C} \} du$. As discussed above, the ratio of the reversal potentials between excitation and inhibition can be viewed as large $(|\frac{\varepsilon_E}{\varepsilon_I}| \gg 1)$, therefore, we have

$$SC \approx -\varepsilon_E \int_0^t \{ \frac{G_I(u)}{C^2} Q(G_E, u) - \frac{\alpha}{C} G_E(u) G_I(u) e^{G_L(u-t)/C} \} du,$$

which is independent of ε_I . Therefore, a moderate change of the driving force for IPSP in the DIF model would not affect the value of SC. As shown in Fig. 6C, the result of our DIF model agrees with the experimental observation. This independence of the inhibitory driving force shows that the nonlinear term in Eq. (1) is

consistent with the notion of shunting inhibition [20]. Using Eq. (2) along with the above equation, we obtain

$$k^{M} = \frac{SC}{V_{E}(t) \cdot V_{I}(t)}$$
$$\approx -\frac{1}{\varepsilon_{I}} \frac{\int_{0}^{t} \{h_{I}(u)Q(h_{E},u) - \alpha Ch_{E}(u)h_{I}(u)e^{G_{L}(u-t)/C}\}du}{Q(h_{E},t)Q(h_{I},t)}$$

which shows that the shunting coefficient k^M is inversely proportional to ε_I in the DIF model. This relation has also been found by using the multi-compartmental model in Ref. [20].

DIF Model for Neuronal Network

Note that a pyramidal neuron normally receives a large number of excitatory and inhibitory synaptic inputs [37]. It has been examined by simulation [20] whether the dendritic integration rule obtained from a pair of excitation and inhibition is applicable to multiple excitatory and inhibitory inputs. By using the NEURON software, the following relationship has been found

$$V_{S} = \sum_{i} V_{E}^{i} + \sum_{j} V_{I}^{j} + \sum_{ij} k_{ij} V_{E}^{i} V_{I}^{j}, \qquad (11)$$

where V_S is the SSP with a coincident activation of all excitatory and inhibitory inputs, V_E^i and V_I^j are the individual EPSP and IPSP, respectively. V_E^i is induced by the *i*th excitatory input alone and V_I^j is induced by the *j*th inhibitory input alone [20]. To account for multiple excitatory and inhibitory inputs and related dendritic integrations, we need to further generalize the DIF model. We propose the following natural extension:

$$C\frac{dV^{l}}{dt} = -G_{L}(V^{l} - \varepsilon_{L}) - \sum_{i} G_{E}^{i}(V^{l} - \varepsilon_{E}) - \sum_{j} G_{I}^{j}(V^{l} - \varepsilon_{I})$$

$$-\sum_{i} \sum_{j} \alpha_{ij}^{l} G_{E}^{i} G_{I}^{j}(V^{l} - \varepsilon_{E}),$$
(12)

where V^l is the membrane potential of the *l*th neuron, G_F^i represents the *i*th excitatory conductance input and G_I^j the *j*th inhibitory conductance input, α_{ij}^l is determined by the shunting coefficient k_{ij} for the pair of the *i*th excitatory and the *j*th inhibitory inputs. Using the above model, we study whether the dendritic integration rule has the form of Eq. (11). We tested the case of two excitatory and two inhibitory inputs using several groups of $\{\alpha_{ii}\}$, with each α_{ii} corresponding to the shunting coefficient for each pair of excitation and inhibition. The results are shown in Fig. 6D, which shows that the form of Eq. (11) holds as the dendritic integration rule in the DIF network model [Eq. (12)]. Similar to Eq. (1), Eq. (11) requires that all excitatory and inhibitory inputs occur simultaneously, therefore, one cannot use the formula [Eq. (11)] to calculate the soma responses to the general stimuli of multiple inputs. To study neuronal responses to general inputs of multiple sites, we need to use the DIF network model since it naturally exhibits dendritic integration effects dynamically.

For a given network of N neurons with polysynaptic connectivity, we can use the value of α_{ij}^l for the *l*th neuron in Eq. (12) to effectively take into account the dendritic integration effect arising from each pair of synaptic inputs from the *i*th presynaptic excitatory neuron and the *j*th presynaptic inhibitory neuron. The values of $\{\alpha_{ij}^l\}$ are chosen to model spatial distances of the synaptic locations. Then, we can evolve the dynamics of the neuronal network [Eq. (12)] without explicitly considering the dendritic tree structure for each neuron. Clearly, neither the polysynaptic connectivity nor the value of α_{ij}^l for each pair of synaptic inputs is easy to obtain in current experiments. However, one may numerically study the network dynamics by choosing different values of $\{\alpha_{ij}^l\}$, which correspond to different spatial effects of dendritic integration. Our DIF network model might be potentially useful in the numerical study to address the effects arising from dendritic integration in neuronal networks.

Discussion

In this work, we have proposed a simple DIF neuron model that dynamically incorporates the spatial dependence of the dendritic integration rule by a phenomenological parametrization. Via analytical and numerical methods, we have shown that the DIF model is capable of capturing many experimental observations. Below, we will further discuss properties of this model as well as predictions of this model.

First, we will provide some rationale to the form of the DIF model in Eq. (10). In fact, Eq. (10) can be viewed as a special case of the following equation

$$C\frac{d}{dt}V = -G_L(V - \varepsilon_L) - G_E f(G_I)(V - \varepsilon_E) - G_I(V - \varepsilon_I), \quad (13)$$

where $f(G_I) = 1 + \alpha_1 G_I(t) + \alpha_2 G_I^2(t) + \cdots$. As we have discussed previously, higher order terms of G_I can be ignored since they are too small to have any significant influence on the value of the shunting coefficient k. Therefore, Eq. (10) essentially encompasses the major terms which contribute to the dendritic integration [Eq. (1)]. The nonlinear SC term in Eq. (1), which takes the form of the product between excitatory and inhibitory responses, can be understood as follows: the input is through conductances which appear as the multiplication factor of the voltage in Eq. (14). Therefore, linear summation rule for the responses is not necessarily true since the relation between the input and the output response is no longer linear. In other words, the bilinear structure between the conductance and the voltage in I&F-type models gives rise to the bilinear term of the excitatory response V_E and the inhibitory response V_I when the excitatory and inhibitory conductance inputs occur simultaneously. Note that, the voltage of neurons in the experiment [20] is substantially below the threshold, therefore, we can use the linear component of the I&F model to model a neuron's dynamical behavior. However, when the voltage is not sufficiently low or when a neuron produces spikes, we may need to use the exponential I&F model [34] or Hodgkin-Huxley-type neuron models to take into account the nonlinear behaviors of a neuron arising from its ion channels. Of course, further experiments should be performed to examine dendritic integration effects in such regimes.

Next, we determine the range of the parameter α in our DIF model. Notice that, for a fixed excitatory input location, the shunting coefficient k measured in the experiment is between 0.08 and 0.3mV^{-1} for various inhibitory input locations [20]. From the relation [Eq. (9)] between k^M and α , we can determine the range of α : $-25 \sim -2 \text{ k}\Omega \cdot \text{cm}^2$. Interestingly, in this range, α is always negative. As a consequence, the inhibitory conductance input will reduce the effects of excitatory drive, as can be seen from the term $G_E(1 + \alpha G_I)(\varepsilon_E - V)$ in Eq. (10). In other words, the inhibition is

Predictions by the DIF Model

We now turn to the prediction of our DIF model, which can be verified in experiments:

(1) The asymptotic behaviors of k^* and k^M are quite different when the time *t* is near zero. As shown in Fig. 6E, k^* approaches a finite value, whereas k^M approaches infinity as *t* tends to zero. From Eq. (9), the difference between k^* and k^M is $\alpha C \int_0^t h_E(u) h_I(u) e^{G_L(u-t)/C} du$

 $\frac{\partial C}{\partial Q} \frac{\int_{0} h_{E}(u) h_{I}(u) e^{-u t} du}{\varepsilon_{I} Q(h_{I}, t) Q(h_{E}, t)}, \text{ which we refer to as the remainder term and its asymptotic behavior is <math>\sim t^{-1}$ when t is near zero. In

term and its asymptotic behavior is $\sim t^{-1}$ when t is hear zero. In this case, although both the numerator and the denominator of the remainder term are small, the ratio between them can still be very large since the numerator is $\sim t^{3}$ and the denominator is $\sim t^{4}$ when t is near zero. By measuring the EPSP, IPSP and SSP at the early time t instead of $t^{*} \approx 20$ ms in the experiment [20], one could first examine whether the dendritic integration rule [Eq. (1)] still holds. If so, one could further verify whether there is an increase in magnitude for the shunting coefficient k as discussed above. Of course, care should be taken in measuring k near t=0 because in this situation the signals are very weak, thus leading to a larger measurement error for k.

(2) As discussed previously, the intrinsic origin of the product between EPSP and IPSP in the dendritic integration rule comes from the product between the excitatory and inhibitory conductances. From the DIF model, it is easy to derive the nonlinear relation $G_{EI} \approx G_L + G_E + G_I + \alpha G_E G_I$, where G_{EI} is the conductance when both excitatory and inhibitory inputs are presented. By recording EPSP, IPSP and SSP with high temporal resolution [34], one can construct the corresponding conductances G_E , G_I and G_{EI} to examine whether such a nonlinear relation holds for any given fixed pair of excitatory and inhibitory inputs.

(3) For synaptic inputs, e.g., inputs are through conductances G_E and G_I , the I&F-type models are no longer linear because they contain the product between the input (conductance) and the response (voltage) as in Eqs. (10) and (14). However, if all the inputs are through direct current injection, the nonlinear SC term in Eq. (1) should vanish as predicted by both the DIF model and the standard I&F model [Eq. (14)].

(4) From the expression of the shunting coefficient k^M [Eq. (8)], we notice that this dendritic integration rule is not completely attributable to the dendritic properties. The first term on the righthand-side of Eq. (8) shows that the somatic membrane properties are also responsible for the integration rule. Therefore, if all the conductance inputs are acted on the soma instead of on the dendrite, one should also observe the dendritic integration rule as in Eq. (1). Of course, in this case, the shunting coefficient k will no longer possess (i) the distal-proximal asymmetry property, and (ii) the dependence of active conductance on the dendrite as observed in the experiment [20] since the input sites of excitation and inhibition are all located at one point (the soma) and the distribution of active conductances (ion channels and receptors) on the dendrite are no longer relevant.

Finally, we point out that the rise and decay time constants of EPSP depend on the input sites for real neurons [38]. Since the shunting coefficient k^* in Eq. (3) is a function of σ_{Er} , σ_{Ed} , one may wonder whether it is possible to capture the spatial dependence of the shunting coefficient k by the I&F model [Eq. (14)] in combination with the input-location dependence of the time constant of EPSP. We first substitute the EPSP's rise and decay time constants measured in the experiment [38] into the fitting

function in Ref. [38] to reconstruct the EPSP profile. Then, we use a differential evolution method [39]—which is a global optimization method—to search for the best choices of σ_{Er} and σ_{Ed} to fit the reconstructed EPSP profile [38]. As shown in Fig. 6F, the value of k^* calculated in this manner first decreases as the distance increases between the excitatory input site and the soma, and then saturates at a constant value. The behavior is not consistent with the experimental measurements [20] (the inset of Fig. 6F). In addition, this approach also fails to explain the phenomena mentioned in Fig. 6B.

Methods

Conductance-based Integrate-and-fire Model

For the conductance-based I&F neuron model, its dynamics are governed by [33]

$$C\frac{dV}{dt} = -G_L(V - \varepsilon_L) - G_E(V - \varepsilon_E) - G_I(V - \varepsilon_I), \quad (14)$$

where *C* is the membrane capacitance per unit area, G_L is the leaky conductance, G_E and G_I are the excitatory and inhibitory conductances, respectively. ε_L is the resting potential, ε_E and ε_I are the excitatory and inhibitory reversal potentials, respectively. The dynamics of conductances G_E and G_I can be described by [40]

$$G_E(t) = f_E N_E h_E(t), \tag{15}$$

$$G_I(t) = f_I N_I h_I(t), \tag{16}$$

where f_E and f_I represent the input strength of excitation and inhibition, respectively. N_E and N_I are normalization factors which make f_E and f_I the maxima of G_E and G_I , respectively. They are chosen as $N_E = [(\frac{\sigma_{Er}}{\sigma_{Ed}})^{\frac{\sigma_{Er}}{\sigma_{Ed}}-\sigma_{Er}} - (\frac{\sigma_{Er}}{\sigma_{Ed}})^{\frac{\sigma_{Ed}}{\sigma_{Ed}}-\sigma_{Er}}]^{-1}$, $N_I = [(\frac{\sigma_{Ir}}{\sigma_{Id}})^{\frac{\sigma_{Ir}}{\sigma_{Id}}-\sigma_{Ir}} - (\frac{\sigma_{Ir}}{\sigma_{Id}})^{\frac{\sigma_{Id}}{\sigma_{Id}}-\sigma_{Ir}}]^{-1}$. σ_{Er} and σ_{Ed} are the rise and decay time constants of the excitatory conductance, respectively. σ_{Ir} and σ_{Id} are the rise and decay time constants of the inhibitory conductance, respectively. $h_E(t)$ and $h_I(t)$ are α -like functions [40] which determine the profiles of EPSP and IPSP, respectively. They are defined as

$$h_E(t) = e^{-\frac{t}{\sigma_{Ed}}} - e^{-\frac{t}{\sigma_{Er}}},\tag{17}$$

$$h_I(t) = e^{-\frac{t}{\sigma_{Id}}} - e^{-\frac{t}{\sigma_{Ir}}}.$$
(18)

Since the I&F model only describes the soma dynamics, G_E and G_I are not synaptic conductances but the effective soma conductances which model the change of the somatic membrane potential due to local synaptic inputs on the dendrite.

In order to be consistent with the setup in the experiment [20], all the voltage values mentioned are chosen as relative voltages with respect to the resting potential. To reproduce the same profiles of EPSP and IPSP as measured in the experiment [20], we choose the time constants as $\sigma_{Er} = 5$ ms, $\sigma_{Ed} = 7.8$ ms, $\sigma_{Ir} = 6$ ms, and $\sigma_{Id} = 18$ ms. The range of input strength f_E and f_I is

determined by the range of amplitudes of EPSP and IPSP in the experiment [20]. f_E is chosen from 1.8×10^{-6} to $1.8 \times 10^{-5} S \cdot cm^{-2}$ to make the EPSP vary from 1 mV to 10 mV. Similarly, f_I is chosen from 1.7×10^{-6} to $5.2 \times 10^{-5} S \cdot cm^{-2}$ to make IPSP vary from 0.2 mV to 4 mV. The reversal potentials relative to the resting potential are chosen to be the same as those used in the experiment [20]: $\varepsilon_L = 0$ mV, $\varepsilon_E = 70$ mV, $\varepsilon_I = -10$ mV, along with commonly used neurophysiological parameters $C = 1.0 \times 10^{-6}$ F·cm⁻², $G_L = 5.0 \times 10^{-5} S \cdot cm^{-2}$ measured in experiments [33]. As shown in Fig. 3, the profiles of EPSP and IPSP measured in the experiment can be well reproduced by the I&F model with the above parameters.

Analytical and Approximate Solutions to I&F-type Models

Based on the I&F model, the individual EPSP $[V_E(t)]$ and IPSP $[V_I(t)]$ under separate excitatory and inhibitory inputs can be described by

$$C\frac{d}{dt}V_E = -G_L(V_E - \varepsilon_L) - G_E(V_E - \varepsilon_E), \qquad (19)$$

$$C\frac{d}{dt}V_I = -G_L(V_I - \varepsilon_L) - G_I(V_I - \varepsilon_I), \qquad (20)$$

whereas, the SSP $[V_S(t)]$ under simultaneous excitatory and inhibitory inputs can be described by

$$C\frac{d}{dt}V_S = -G_L(V_S - \varepsilon_L) - G_E(V_S - \varepsilon_E) - G_I(V_S - \varepsilon_I). \quad (21)$$

The conductances of excitation and inhibition are given by Eqs. (15) and (16). With notations $G_S = G_E + G_I$ and $\widetilde{G}_S = \varepsilon_E G_E + \varepsilon_I G_I$, we can obtain analytical solutions to Eqs. (19–21) along with their approximations in integral forms as

$$V_E(t) = \int_0^t \varepsilon_E \frac{G_E(u)}{C} e^{G_L(u-t)/C} e^{\int_t^u G_E(v)/Cdv} du$$
(22a)

$$\approx \int_0^t \varepsilon_E \frac{G_E(u)}{C} e^{G_L(u-t)/C} (1 + \int_t^u G_E(v)/Cdv) du, \quad (22b)$$

$$V_I(t) = \int_0^t \varepsilon_I \frac{G_I(u)}{C} e^{G_L(u-t)/C} e^{\int_t^u G_I(v)/Cdv} du$$
(23a)

$$\approx \int_0^t \varepsilon_I \frac{G_I(u)}{C} e^{G_L(u-t)/C} (1 + \int_t^u G_I(v)/Cdv) du, \quad (23b)$$

$$V_{\mathcal{S}}(t) = \int_0^t \frac{\widetilde{G}_{\mathcal{S}}(u)}{C} e^{G_L(u-t)/C} e^{\int_t^u G_{\mathcal{S}}(v)/Cdv} du$$
(24a)

$$\approx \int_0^t \frac{\widetilde{G}_{\mathcal{S}}(u)}{C} e^{G_L(u-t)/C} (1 + \int_t^u G_{\mathcal{S}}(v)/Cdv) du, \qquad (24b)$$

where approximations are taken with respect to the second order

of $\int_{t}^{u} G_{E}(s)ds$, $\int_{t}^{u} G_{I}(s)ds$, and $\int_{t}^{u} G_{S}(s)ds$ in Taylor expansions. In particular, the soma response $V_{E}(t)$ under only the excitatory input can also be approximated by the first-order expansion as $V_{E}(t) \approx \int_{0}^{t} \varepsilon_{E} \frac{G_{E}(u)}{C} e^{G_{L}(u-t)/C} du.$

For the modified I&F model [Eq. (7)], the individual EPSP (V_E^M) can be obtained by setting the inhibitory input $G_I = 0$ in Eq. (7). For this case, Eq. (7) reduces to Eq. (19). Similarly, for the individual IPSP (V_I^M) where $G_E = 0$, Eq. (7) reduces to Eq. (20). Therefore, we can use Eqs. (23) and (25) as approximations of EPSP (V_E^M) and IPSP (V_I^M) . For the SSP (V_S^M) , with notations $G_S^M = G_E + G_I + (\alpha + \beta)G_EG_I, \ \widetilde{G_S^M} = \varepsilon_E G_E + \varepsilon_I G_I + (\alpha \varepsilon_E + \beta \varepsilon_I)G_EG_I$, we can obtain the analytical solution to Eq. (7) as

$$V_{S}^{M}(t) = \int_{0}^{t} \frac{\widetilde{G_{S}^{M}}(u)}{C} e^{G_{L}(u-t)/C} e^{\int_{t}^{u} G_{S}^{M}(v)/Cdv} du$$
(25)

along with the following approximation by performing the secondorder Taylor expansion with respect to $\int_{-1}^{u} G_{S}^{M}(v) dv$:

$$V_S^M(t) \approx V_S(t) + \left(\frac{\alpha}{C}\varepsilon_E + \frac{\beta}{C}\varepsilon_I\right) \int_0^t G_E(u)G_I(u)e^{G_L(u-t)/C}du, \quad (26)$$

where $V_S(t)$ is given by Eq. (24b). All the above approximations have been verified numerically and the relative errors with respect to the analytical solutions are less than 5%.

Two-port Analysis

A linear relationship between the synaptic current and the membrane potential has been observed in the experiment [41] for fast, non-NMDA input into hippocampal pyramidal neurons. Therefore, we can describe the synaptic currents of excitation and inhibition by $I_{ex} = G_E(t)(\varepsilon_E - V_E^E(t))$ and $I_{in}(t) = G_I(t)(\varepsilon_I - V_E^E(t))$ $V_I^I(t)$, respectively. Here, $V_E^E(t)$ and $V_I^I(t)$ represent the membrane potentials at the sites of excitation and inhibition, respectively. According to linear cable theory [33], the voltage change $V_i(t)$ at location j in response to an arbitrary current input $I_i(t)$ at location i can be expressed as $V_i(t) = K_{ii}(t) \star I_i(t)$, where $K_{ii}(t)$ is the impulse response of the system and the symbol " \star " stands for convolution in time. In particular, for the timeindependent case, we can obtain the EPSP at the soma (V_E) under only the excitatory input as $V_E = K_{ES}G_E(\varepsilon_E - V_E^E)$, whereas the EPSP at the input site (V_E^E) can be obtained as $V_E^E = K_{EE}G_E(\varepsilon_E - V_E^E)$. Therefore, we have

$$V_E = \frac{K_{ES} G_E \varepsilon_E}{1 + K_{EE} G_E}.$$
(27)

Similarly, we can obtain the IPSP at the soma (V_I) under only the inhibitory input as

$$V_I = \frac{K_{IS}G_I\varepsilon_I}{1 + K_{II}G_I}.$$
(28)

For simultaneous inputs of excitation and inhibition, the SSP can be expressed as

$$V_{S} = K_{ES}G_{E}(\varepsilon_{E} - \widetilde{V_{E}}) + K_{IS}G_{I}(\varepsilon_{I} - \widetilde{V_{I}}), \qquad (29)$$

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where $\widetilde{V_E}$ and $\widetilde{V_I}$ are given by $\widetilde{V_E} = K_{EE}G_E(\varepsilon_E - \widetilde{V_E}) + K_{IE}G_I(\varepsilon_I - \widetilde{V_I})$ and $\widetilde{V_I} = K_{II}G_I(\varepsilon_I - \widetilde{V_I}) + K_{EI}G_E(\varepsilon_E - \widetilde{V_E})$, respectively. Solving the above equations, we can obtain

$$\widetilde{V}_E = \frac{K_{EE}G_E\varepsilon_E + K_{IE}G_I\varepsilon_I + (K_{EE}K_{II} - K_{EI}K_{IE})G_EG_I\varepsilon_E}{1 + K_{EE}G_E + K_{II}G_I + (K_{EE}K_{II} - K_{EI}K_{IE})G_EG_I}, \quad (30)$$

 $\widetilde{V}_I =$

$$-\frac{(K_{EE}G_{E}\varepsilon_{E}+K_{IE}G_{I}\varepsilon_{I})K_{EI}G_{E}+(K_{EI}G_{E}\varepsilon_{E}+K_{II}G_{I}\varepsilon_{I})(1+K_{EE}G_{E})}{1+K_{EE}G_{E}+K_{II}G_{I}+(K_{EE}K_{II}-K_{EI}K_{IE})G_{E}G_{I}},$$
(31)

 $V_S =$

$$\frac{K_{ES}G_E\varepsilon_E + K_{IS}G_I\varepsilon_I + (K_{ES}K_{II} - K_{IS}K_{EI})G_IG_E\varepsilon_E + (K_{IS}K_{EE} - K_{ES}K_{IE})G_EG_I\varepsilon_I}{1 + K_{EE}G_E + K_{II}G_I + (K_{EE}K_{II} - K_{EI}K_{IE})G_EG_I}.$$
(32)

As pointed out in Refs. [20,33], the magnitudes of $K_{EE}G_E$, $K_{II}G_I$, $K_{IE}G_I$, $K_{EI}G_E$ are on the order of $10^{-2} \sim 10^{-1}$ due to small synaptic inputs and thus can be viewed as much smaller than unity. Therefore, we have the following approximations by keeping up to the second-order terms in Taylor expansions

$$V_E \approx K_{ES}(1 - K_{EE}G_E)G_E\varepsilon_E, \tag{33}$$

References

- Rall W, Segev I, Rinzel J, Shepherd G (1995) The theoretical foundation of dendritic function. Cambridge: MIT press.
- Rall W (1964) Theoretical significance of dendritic trees for neuronal inputoutput relations. In Neural Theory and Modeling, ed. R.F. Reiss, Stanford University Press, Stanford CA.
- Burke R (1967) Composite nature of the monosynaptic excitatory postsynaptic potential. J Neurophysiol 30: 1114–1137.
- Kuno M, Miyahara J (1969) Non-linear summation of unit synaptic potentials in spinal motoneurones of the cat. J Physiol 201: 465.
- Schwindt P, Crill W (1998) Synaptically evoked dendritic action potentials in rat neocortical pyramidal neurons. J Neurophysiol 79: 2432–2446.
- Margulis M, Tang C (1998) Temporal integration can readily switch between sublinear and supralinear summation. J Neurophysiol 79: 2809–2813.
- 7. Nettleton J, Spain W (2000) Linear to supralinear summation of ampa-mediated epsps in neocortical pyramidal neurons. J Neurophysiol 83: 3310–3322.
- Poirazi P, Brannon T, Mel B (2003) Arithmetic of subthreshold synaptic summation in a model cal pyramidal cell. Neuron 37: 977–987.
- Poirazi P, Brannon T, Mel B (2003) Pyramidal neuron as two-layer neural network. Neuron 37: 989–999.
- Polsky A, Mel B, Schiller J (2004) Computational subunits in thin dendrites of pyramidal cells. Nat Neurosci 7: 621–627.
- Losonczy A, Magee J (2006) Integrative properties of radial oblique dendrites in hippocampal cal pyramidal neurons. Neuron 50: 291–307.
- Häusser M, Mel B (2003) Dendrites: bug or feature? Curr Opin Neurobiol 13: 372–383.
- London M, Häusser M (2005) Dendritic computation. Annu Rev Neurosci 28: 503–532.
- David F, Linster C, Cleland T (2008) Lateral dendritic shunt inhibition can regularize mitral cell spike patterning. J Comput Neurosci 25: 25–38.
- Ariel M, Kogo N (2005) Shunting inhibition in accessory optic system neurons. J Neurophysiol 93: 1959–1969.
- Chacron M (2006) Nonlinear information processing in a model sensory system. J Neurophysiol 95: 2933–2946.
- Mitchell S, Silver R (2003) Shunting inhibition modulates neuronal gain during synaptic excitation. Neuron 38: 433–445.
- Atallah B, Scanziani M (2009) Instantaneous modulation of gamma oscillation frequency by balancing excitation with inhibition. Neuron 62: 566–577.
- Vida I, Bartos M, Jonas P (2006) Shunting inhibition improves robustness of gamma oscillations in hippocampal interneuron networks by homogenizing firing rates. Neuron 49: 107–117.
- Hao J, Wang X, Dan Y, Poo M, Zhang X (2009) An arithmetic rule for spatial summation of excitatory and inhibitory inputs in pyramidal neurons. Proc Natl Acad Sci USA 106: 21906–21911.
- 21. Carnevale N, Hines M (2006) The NEURON book. Cambridge: Cambridge University Press.

 $V_I \approx K_{IS} (1 - K_{II} G_I) G_I \varepsilon_I, \tag{34}$

and

$$V_{S} \approx K_{ES}(1 - K_{EE}G_{E})G_{E}\varepsilon_{E} + K_{IS}(1 - K_{II}G_{I})G_{I}\varepsilon_{I}$$

$$- K_{ES}K_{IE}G_{E}G_{I}\varepsilon_{I} - K_{IS}K_{EI}G_{E}G_{I}\varepsilon_{E}.$$
(35)

Finally, we can obtain

$$V_S \approx V_E + V_I - \left(\frac{K_{IE}}{K_{IS}\varepsilon_E} + \frac{K_{EI}}{K_{ES}\varepsilon_I}\right) V_E V_I \tag{36}$$

with the shunting coefficient $k = -\frac{K_{IE}}{K_{IS}\varepsilon_E} - \frac{K_{EI}}{K_{ES}\varepsilon_I}$. Eq. (36) has the same form as the dendritic integration rule [Eq. (1)] as observed in the experiment.

Author Contributions

Conceived and designed the experiments: DZ SL XhZ DC. Performed the experiments: DZ SL XhZ DC. Analyzed the data: DZ SL XhZ DC. Contributed reagents/materials/analysis tools: DZ SL XhZ DC. Wrote the paper: DZ SL XhZ DC.

- Wehr M, Zador A (2003) Balanced inhibition underlies tuning and sharpens spike timing in auditory cortex. Nature 426: 442–446.
- Lapique L (1907) Recherches quantitatives sur l'excitation electrique des nerfs traitee comme une polarization. J Physiol Pathol Gen 9: 620–635.
- Smith G, Cox C, Sherman S, Rinzel J (2000) Fourier analysis of sinusoidally driven thalamocortical relay neurons and a minimal integrate-and-fire-or-burst model. J Neurophysiol 83: 588–610.
- Hodgkin A, Huxley A (1952) A quantitative description of membrane current and its application to conduction and excitation in nerve. J Physiol 117: 500– 544.
- Izhikevich E (2003) Simple model of spiking neurons. IEEE Trans Neural Netw 14: 1569–1572.
- Fitzhugh R (1961) Impulses and physiological states in theoretical models of nerve membrane. Biophys J 1: 445–466.
- Rose R, Hindmarsh J (1989) The assembly of ionic currents in a thalamic neuron i. the three-dimensional model. Proc R Soc Lond B Biol Sci 237: 267– 288.
- Morris C, Lecar H (1981) Voltage oscillations in the barnacle giant muscle fiber. Biophys J 35: 193–213.
- Wilson H (1999) Simplified dynamics of human and mammalian neocortical neurons. J Theor Biol 200: 375–388.
- Izhikevich E (2004) Which model to use for cortical spiking neurons? IEEE Trans Neural Netw 15: 1063–1070.
- Feng J (2001) Is the integrate-and-fire model good enough?-a review. Neural Netw 14: 955–975.
- Koch C (2005) Biophysics of computation: information processing in single neurons. Oxford: Oxford University Press.
- Badel L, Lefort S, Berger T, Petersen C, Gerstner W, et al. (2008) Extracting non-linear integrate-and-fire models from experimental data using dynamic i–v curves. Biol Cybern 99: 361–370.
- Carandini M, Mechler F, Leonard C, Movshon J, et al. (1996) Spike train encoding by regular-spiking cells of the visual cortex. J Neurophysiol 76: 3425– 3441.
- Cash S, Yuste R (1998) Input summation by cultured pyramidal neurons is linear and position-independent. J Neurosci 18: 10–15.
- Bruno R, Sakmann B (2006) Cortex is driven by weak but synchronously active thalamocortical synapses. Science 312: 1622–1627.
- Magee J, Cook E, et al. (2000) Somatic epsp amplitude is independent of synapse location in hippocampal pyramidal neurons. Nat Neurosci 3: 895–903.
- Storn R, Price K (1997) Differential evolution-a simple and efficient heuristic for global optimization over continuous spaces. J Global Optim 11: 341–359.
- 40. Dayan P, Abbott L (2001) Theoretical neuroscience: Computational and mathematical modeling of neural systems. Cambridge: MIT Press.
- Hestrin S, Nicoll R, Perkel D, Sah P (1990) Analysis of excitatory synaptic action in pyramidal cells using whole-cell recording from rat hippocampal slices. J Physiol 422: 203–225.